

LETTER TO THE EDITOR

Open Access

# PARP inhibitors: current status and implications for anticancer therapeutics

Hadi Usmani<sup>1</sup>, Syed Ather Hussain<sup>1</sup> and Asfandyar Sheikh<sup>1,2\*</sup>

## Abstract

Poly(ADP-ribose) polymerases (PARPs) comprise of a large family of 17 proteins encoded by various genes which participate in genome maintenance, apoptosis, inflammatory responses and the regulation of gene expression programs. PARP inhibitors, as therapeutic agents, come into play acting on both PARP 1 and PARP 2. These drugs seem to target tumor cells in a moment of vulnerability when they are undergoing DNA repair. In the past few years this class of anti-cancer drug has been discovered to show a promising niche in the clinic.

## Letter to the Editor

Poly(ADP-ribose) polymerases (PARPs, aka diphtheria toxin-like ADP-ribosyltransferases [ARTDs]) comprise of a large family of 17 proteins encoded by various genes which participate in genome maintenance, apoptosis, inflammatory responses and the regulation of gene expression programs [1,2]. Out of these 17 proteins, PARP1 and PARP2 are the only enzymes whose catalytic activity is turned on when there is a single strand break (SSB) in the DNA [3].

PARPs constitutes of four functional domains: a DNA-binding domain, a B domain, an auto-modification domain and a catalytic domain. The DNA binding domain comprises of two zinc fingers and is involved in detecting and resealing DNA breaks [4]. In addition to containing a site which is acted upon by caspases resulting in inactivation of PARP, domain B also has a bipartite nuclear localization signal which is involved in the nuclear homing of PARP1 [5]. The automodification domain has a BRCT motif and is responsible for letting go of the protein from DNA after the catalytic reaction has occurred at the catalytic domain. The catalytic domain is involved in the synthesis of the poly (ADP-ribose) chain.

Following DNA damage, PARP binds to nicotinamide adenine dinucleotide and splits it off at the nicotinamide-ribose bond with polymerization of the ADP-ribose moiety [6]. Repetition of this step results in the

synthesis of a multi-branched polymer of ADP-ribose on either nuclear protein acceptors located on chromatin or on the PARP itself [7]. These automodification and heteromodification reactions ensure the longevity of damaged replicating cells because the poly (ADP-ribose) chain attracts the attention of DNA repairing enzymes and scaffolding proteins such as DNA ligase III, DNA polymerase beta and X-ray cross-complementing gene 1 which in turn take part in a process called base excision repair (BER) [8].

Two theories have been proposed by which PARP is thought to induce apoptosis in cells. First, there is the caspase independent pathway in which PARP is excessively stimulated, and the other is the caspase dependent pathway in which PARP is inactivated. In the first pathway, it is believed that PARP1 activation and the subsequent synthesis of PAR polymer serves as a cell death signal which leads to the release of apoptosis-inducing factor (AIF) from the mitochondria into the nucleus [9]. AIF then initiates apoptosis. However, in the other pathway, PARP is rapidly and specifically cleaved by caspase-3, a mammalian ICE-related protease. It is believed that PARP cleavage ensures that all cells undergoing self-destruction turn off protective mechanisms like DNA repair associated with PARP activity [10]. The exact mechanism of these two processes and the intermediate steps involved are not fully understood.

Even without complete apprehension of the mechanisms of the Poly(ADP-ribose) polymerases, there has been significant research towards targeting these proteins in cancer patients. This is where the class of PARP

\* Correspondence: asfandyarsheikh@gmail.com

<sup>1</sup>Dow Medical College, Dow University of Health Sciences, Karachi, Pakistan

<sup>2</sup>Pakistan Research Evolution Scientific Society, Baba-e-Urdu Road, Karachi 74200, Pakistan

inhibitors as therapeutic agents come into play acting on both PARP 1 and PARP 2. These drugs seem to target tumor cells in a moment of vulnerability when they are undergoing DNA repair [11].

Early clinical phase 1 trials were conducted on breast, ovarian and prostate cancer targeting the BRCA1/2 mutations. The rationale for use of PARP inhibitors in these cancers represent an exquisite example of how different pathways cooperate to repair damage [12,13]. BRCA1 and BRCA2 have an essential role in DNA double-strand break repair via homologous recombination, whereas PARP, as mentioned earlier, has a role in BER. It has been shown that BRCA1 or BRCA2 dysfunction sensitizes cells to the PARP inhibition, which subsequently results in chromosomal instability and cell cycle arrest and apoptosis, thereby working synergistically and providing a novel therapeutic option without the need for an exogenous DNA-damaging agent [12,13].

Iniparib (BSI 201) being one of the initial candidates showed serious potential but failed to meet expectations in phase III trials and was eventually disproved as a PARP inhibitor [1]. Olaparib (AZD2281), another PARP inhibitor, produced positive anti-tumor results in cancer patients with BRCA1/2 mutations [14]. Currently there are 8 molecules in clinical trials which follow two distinct pathways of pharmacodynamics. The first is a monotherapy which targets tumor cells that have a defect in the DNA repair mechanisms. The second is a combination therapy with DNA damaging chemotherapeutic treatments thus offering a potentiating effect to cause the cell to undergo apoptosis [1]. Patients evaluated in clinical trials with germline BRCA1/2 mutations, through one of those pathways, have seen a 40% response rate with olaparib in platinum sensitive ovarian cancer [15]. PARPis have also been proven to be the first treatment for BRCA1/2 mutation carrier in prostate cancer patients and have promising anti-tumor activity [16].

It is important to know that BRCA1/2 associated cancers are quite rare and only account for less than 5% of breast-cancer patients so it is crucial to see how PARPis function on other cancers. An interesting match is triple negative breast cancer in which tumors lack expression of the estrogen receptor (ER), progesterone receptor (PR), and human epidermal growth factor receptor 2 (HER2) [11]. In a trial conducted by O'Shaughnessy, triple negative breast cancer patients were given iniparib in conjunction with gemcitabine and carboplatin (chemotherapy doublet) which improved the rate of clinical benefit from 34% to 56% ( $P = 0.01$ ) and the rate of overall response from 32% to 52% ( $P = 0.02$ ) [17]. Unfortunately despite recent advances in the clinical evaluation of various PARP inhibitors in triple-negative breast cancer (TNBC)

patients, there has been very little anti-tumor progression beyond PARPis [18]. Another example is a trial conducted with rucaparib (PF-01367338, AG014699) in combination with temozolomide for the treatment of metastatic melanoma which showed positive signs of chemopotential. Results showed that response rate was 17.4%, median time to progression 3.5 months, median overall survival was 9.9 months, and 36% of patients were progression-free at 6 months [19].

Since PARP inhibitors are blocking the pathway to repair damaged DNA; they require a selective increase in DNA damage to tumor cells compared to normal cells in order to produce an enhanced therapeutic effect by chemotherapy. Thus it would be beneficial if the tumor cells already harbored DNA repair defects such as the case with BRCA1/2 mutations. By blocking both DNA damage repair (DDR) pathways, the first being a tumor germline defect in a non-BER pathway and the second being a PARP inhibited BER pathway, it decreases the chances of the tumor cells to withstand DNA damage. This phenomenon is termed 'synthetic lethality' and is very effective in preventing the tumor cells from replicating efficiently. This theory could also be applied to include treatment of tumors with defects in other HR pathway proteins such as PTEN-deficient cells which are sensitive to PARP inhibitors probably due to the involvement in the expression of RAD51, another gene encoding for DNA double strand break repairs [15].

One thing to note is the limitations of PARP inhibitors with regards to 'PARP trapping' which occurs during PARP inhibitor treatments. In normal cells the PARP1/2 bind to DNA and then recruit other proteins to do the repairing after which the PARPs are released off the DNA. But the inhibitors trap the PARP in the PARP-DNA complex which makes a lethal PARP poison, so to speak. This phenomenon is more detrimental to the cell than the DNA breaks that occur during normal cell replication. Thus PARP inhibitors should be classified not only in their enzyme inhibition abilities but also according to their potency to trap PARP [20].

In the past few years this class of anti-cancer drug has been discovered to show a promising niche in the clinic. There are some limitations including resistance but that is to be expected with cancer cells [21]. But the amount of research being poured into this exciting new class of anti-tumor therapeutics is bound to produce interesting results. Current researchers should be asking the questions related to development of PARP inhibitors that will enhance DNA damage in tumor cells that lack an intrinsic defect in DDR thus being able to divert the synthetic lethality condition [15]. Hopefully with some answers there will be an improved therapy for patients with cancers that have previously been lacking in treatments.

#### Competing interests

The authors declare that no competing interests exist.

#### Authors' contributions

AS conceived the topic. HU and SAH were involved in drafting the initial manuscript. AS critically revised the manuscript. The authors have read and approved the manuscript.

Received: 2 August 2013 Accepted: 30 October 2013

Published: 2 December 2013

#### References

1. Papeo G, Casale E, Montagnoli A, Cirila A: **PARP inhibitors in cancer therapy: an update.** *Expert Opin Ther Pat* 2013, **23**(4):503–514.
2. Kim MY, Zhang T, Kraus WL: **Poly(ADP-ribosylation) by PARP-1: 'PAR-laying' NAD<sup>+</sup> into a nuclear signal.** *Genes Dev* 2005, **19**(17):1951–1967.
3. Tentori L, Graziani G: **Chemopotentiality by PARP inhibitors in cancer therapy.** *Pharmacol Res* 2005, **52**(1):25–33.
4. Petrucco S: **Sensing DNA damage by PARP-like fingers.** *Nucleic Acids Res* 2003, **31**(23):6689–6699.
5. Kaufmann SH, Desnoyers S, Ottaviano Y, Davidson NE, Poirier GG: **Specific proteolytic cleavage of poly(ADP-ribose) polymerase: an early marker of chemotherapy-induced apoptosis.** *Cancer Res* 1993, **53**(17):3976–3985.
6. Curtin NJ: **PARP inhibitors and cancer therapy.** In *Poly (ADP-ribosyl) ation*. United States: Springer; 2006:218–233.
7. D'Amours D, Desnoyers S, D'Silva I, Poirier GG: **Poly(ADP-ribosylation) reactions in the regulation of nuclear functions.** *Biochem J* 1999, **342**(Pt 2):249–268.
8. Ame JC, Spenlehauer C, de Murcia G: **The PARP superfamily.** *Bioelectromagnetics* 2004, **26**(8):882–893.
9. Yu SW, Andrabi SA, Wang H, Kim NS, Poirier GG, Dawson TM, Dawson VL: **Apoptosis-inducing factor mediates poly(ADP-ribose) (PAR) polymer-induced cell death.** *Proc Natl Acad Sci USA* 2006, **103**(48):18314–18319.
10. Wang ZQ, Stingl L, Morrison C, Jantsch M, Los M, Schulze-Osthoff K, Wagner EF: **PARP is important for genomic stability but dispensable in apoptosis.** *Genes Dev* 1997, **11**(18):2347–2358.
11. Carey LA, Sharpless NE: **PARP and cancer—if it's broke, don't fix it.** *N Engl J Med* 2011, **364**(3):277–279.
12. Farmer H, McCabe N, Lord CJ, Tutt AN, Johnson DA, Richardson TB, Santarosa M, Dillon KJ, Hickson I, Knights C, *et al*: **Targeting the DNA repair defect in BRCA mutant cells as a therapeutic strategy.** *Nature* 2005, **434**(7035):917–921.
13. Bryant HE, Schultz N, Thomas HD, Parker KM, Flower D, Lopez E, Kyle S, Meuth M, Curtin NJ, Helleday T: **Specific killing of BRCA2-deficient tumours with inhibitors of poly(ADP-ribose) polymerase.** *Nature* 2005, **434**(7035):913–917.
14. Fong PC, Boss DS, Yap TA, Tutt A, Wu P, Mergui-Roelvink M, Mortimer P, Swaisland H, Lau A, O'Connor MJ, *et al*: **Inhibition of poly(ADP-ribose) polymerase in tumors from BRCA mutation carriers.** *N Engl J Med* 2009, **361**(2):123–134.
15. Kummar S, Chen A, Parchment RE, Kinders RJ, Ji J, Tomaszewski JE, Doroshow JH: **Advances in using PARP inhibitors to treat cancer.** *BMC Med* 2012, **10**:25.
16. Sandhu SK, Omlin A, Hylands L, Miranda S, Barber LJ, Riisnaes R, Reid AH, Attard G, Chen L, Kozarewa I, *et al*: **Poly (ADP-ribose) polymerase (PARP) inhibitors for the treatment of advanced germline BRCA2 mutant prostate cancer.** *Ann Oncol* 2013, **24**(5):1416–1418.
17. O'Shaughnessy J, Osborne C, Pippen JE, Yoffe M, Patt D, Rocha C, Koo IC, Sherman BM, Bradley C: **Iniparib plus chemotherapy in metastatic triple-negative breast cancer.** *N Engl J Med* 2011, **364**(3):205–214.
18. Chuang HC, Kapuriya N, Kulp SK, Chen CS, Shapiro CL: **Differential anti-proliferative activities of poly(ADP-ribose) polymerase (PARP) inhibitors in triple-negative breast cancer cells.** *Breast Cancer Res Treat* 2012, **134**(2):649–659.
19. Plummer R, Lorigan P, Steven N, Scott L, Middleton MR, Wilson RH, Mulligan E, Curtin N, Wang D, Dewji R, *et al*: **A phase II study of the potent PARP inhibitor, rucaparib (PF-01367338, AGO14699), with temozolomide in patients with metastatic melanoma demonstrating evidence of chemopotentiality.** *Cancer Chemother Pharmacol* 2013, **71**(5):1191–1199.
20. Murai J, Huang SY, Das BB, Renaud A, Zhang Y, Doroshow JH, Ji J, Takeda S, Pommier Y: **Trapping of PARP1 and PARP2 by clinical PARP inhibitors.** *Cancer Res* 2012, **72**(21):5588–5599.
21. Fojo T, Bates S: **Mechanisms of resistance to PARP inhibitors—three and counting.** *Cancer Discov* 2013, **3**(1):20–23.

doi:10.1186/1750-9378-8-46

**Cite this article as:** Usmani *et al*: **PARP inhibitors: current status and implications for anticancer therapeutics.** *Infectious Agents and Cancer* 2013 **8**:46.

**Submit your next manuscript to BioMed Central and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
www.biomedcentral.com/submit

