## RESEARCH

## **Open Access**



# The prognostic role of PD-L1 expression and the presence of polyomavirus in Merkel cell carcinoma cases

Stella Meireles Sigueira<sup>1\*</sup>, Gabriella Campos-do-Carmo<sup>2</sup>, Paulo Ricardo Garcia da Silva<sup>3</sup>, Isabele Ávila Small<sup>1</sup><sup>1</sup><sup>1</sup> and Andreia Cristina De Melo<sup>1</sup>

## Abstract

Background Merkel cell carcinoma (MCC) comprises a rare malignant primary skin tumor presenting neuroendocrine differentiation. Recently, agents blocking the programmed cell death protein 1 and programmed cell death protein ligand 1 pathway (PD-1/PD-L1) have demonstrated objective and durable tumor regressions in patients presenting advanced MCC. This study aimed to describe the sociodemographic, clinical, and histopathological characteristics of MCC patients, also assessing the prevalence of PD-L1 expression and Merkel cell Polyomavirus (MCPyV), as well as their prognostic roles.

Methods Data from patients diagnosed with MCC between 1996 and 2019 at a reference cancer center in Rio de Janeiro, southeastern Brazil, were evaluated in a retrospective study. Tumor samples were tested for MCPyV and PD-L1 employing immunohistochemistry. Survival analyses were carried out employing the Kaplan-Meier method and curves were compared using the log-rank test. A multiple semiparametric Cox model was used. Values p < 0.05were considered significant.

**Results** A total of 65 patients were included in the study, with a mean age at diagnosis of 72 (standard deviation 13.9). A total of 56.9% (37/65) of the patients were male, 86.2% (56/65) were white, and 56.9% (37/64) were illiterate or with incomplete elementary school. MCPyV immunohistochemistry was positive in 29 cases (44.6%) and PD-L1 positivity was  $\geq$  1% in 42 cases (64.6%). Significant associations between MCPyV and PD-L1 expression  $\geq$  1% (p = 0.003) and PD-L1 expression  $\ge$  5% (p = 0.005) were noted. Concerning the multivariate analysis, only education level and advanced MCC stage indicated statistically significant worse progression-free survival. Regarding overall survival (OS), being male, education level and advanced stage comprised risk factors. The estimated OS at 60 months for stages I to III was of 48.9% and for stage IV, 8.9%.

Conclusions This is the first large Brazilian cohort to assess the prevalence of MCPyV in MCC tumors, as well as PD-L1 expression and their associations. No correlations were noted between MCPyV infection or PD-L1 expression and survival rates.

Keywords Merkel cell carcinoma, Merkel cell polyomavirus, Programmed death ligand-1, Skin cancer

\*Correspondence: Stella Meireles Sigueira stellamsigueira@gmail.com Full list of author information is available at the end of the article



© The Author(s) 2023. Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

## Background

Merkel cell carcinoma (MCC) is a rare and aggressive type of skin cancer of neuroendocrine origin [1], first described in 1972 by Toker as trabecular skin carcinoma [2]. Since then, knowledge concerning the pathophysiology of this condition and patient management has advanced exponentially. Also known as primary cutaneous neuroendocrine carcinoma, MCC received this denomination due to its ultrastructural and immunophenotypic similarity to Merkel sensory skin cells [2]. However, the Merkel cell origin for MCC was considered unlikely as this carcinoma can actually derive from a neuronal precursor [3–5].

In 2008, Feng et al. described the association between a new polyomavirus and MCC for the first time, later named Merkel cell polyomavirus (MCPyV) [6]. The prevalence of subclinical MCPyV infections, reported as around 60 to 80% in adults, increases with age. This may, however, differ significantly among different geographic regions, such as in Australia, where a much lower association with this viral infection, of around 25%, has been noted [7].

Two distinct MCC etiologies have been described: clonal integration of MCPyV DNA into tumor genomes and UV damage. Both forms exhibit high proliferative growth rates due to mutations in oncogenes and tumor suppressor genes. These include the RB, p53, MYCL, and ATOH1 pathways [8]. Furthermore, emerging evidence indicates that dysregulation of epigenetic mechanisms, including histone post-translational modifications, are involved in MCC. Furthermore, histone acetylation, methylation and phosphorylation mark impairments, as well as histone modifying enzymes, have also been implicated in this disease [9].

Still concerning MCC causes, chronic immunosuppression, including use of immunosuppressive drugs, HIV/ AIDS, lymphoproliferative disorders, solid organ transplantation and auto-immune diseases have all been associated with increased risk of developing MCC. Compared to immune-competent MCC patients, immunosuppressed MCC patients display significantly worse MCCspecific and overall survival [10].

Results are variable regarding the prognostic role of MCPyV status, with predominantly worse prognoses observed in patients presenting polyomavirus-negative tumors or with no difference concerning MCPyV-positive tumors [11]. Both MCPyV positive and negative patients can, however, present clinically aggressive and fatal courses [7].

MCC is characteristically aggressive and locally invasive, with high local recurrence rates and the involvement of regional lymph nodes [1]. A recent study reported a 40% recurrence rate at 5 years for this condition, with a first year risk of recurrence of 11% for stage I patients, 33% for stage IIA/IIB, 30% for stage IIIA, 45% for stage IIIB, and 58% for stage IV, with 95% of all recurrences ensuing in the first 3 years. Other risk factors associated with increased recurrence rates comprise immunosuppression, being male, clinically detectable nodal disease, and advanced age [12].

The limited number of therapeutic MCC treatment options leads to an urgent need to determine tumor-specific pathways and possible therapeutic targets [1]. In this sense, an increasing body of evidence on the role of the immune system in MCC control has paved the way for the use of checkpoint inhibitors, namely anti-PD-1 (programmed cell death 1) and anti-PD-L1 (programmed cell death protein ligand 1) [13].

This study, therefore, aimed to assess clinical MCC patient characteristics, the prevalence of MCPyV and PD-L1 expression, as well as the prognostic role of these biomarkers.

## Methods

#### Study design, patient selection, and data collection

This study was approved by the Brazilian National Cancer Institute (INCA) Ethics in Human Research Committee and conducted following Good Clinical Practice guidelines.

INCA patients were included when older than 18 years old and wen presenting a confirmed histopathological MCC diagnosis between 1996 and 2019. Exclusion criteria comprised patients whose records lacked the clinical data of interest and paraffin-embedded tumor samples.

Patient characteristics, comprising age at diagnosis, sex, phototype, comorbidities, staging, related treatments, response, and survival information, were retrospectively obtained from hospital records and entered into the Research Data Capture (REDCap<sup>®</sup>) system. All cases were reclassified following the American Joint Committee on Cancer AJCC (AJCC) Cancer Staging Manual 8th edition [14].

## Histopathological and immunohistochemistry evaluations

A trained pathologist reviewed all specimens concerning the following morphological variables: mitotic index, fibroplasia, inflammatory infiltrate, tumor thickness, ulceration, necrosis, hemorrhage and subtype [15]. Primary tumor samples were preferentially used. If not available, metastases were analyzed.

MCPyV (clone CM2B4, Santa Cruz, 200  $\mu$ g/mL dilution, 0.1 mL) expression was classified as positive when tumor cells exhibited a dark staining. PD-L1 (clone SP263, Ventana) was quantified based on the percentage of positive cells in 10 fields at 10 magnifications over the total cell number. Tumor cell staining was compared

with positive and negative controls. Two cutoff points were employed for the PD-L1 evaluations, namely <1% (negative) and  $\geq$  1% (positive) or <5% (negative) and  $\geq$  5% (positive).

#### Statistical analyses

Statistical analyses were performed in the R environment [16]. A descriptive analysis of the investigated variables was performed. Means and standard deviations (SD) (or medians and interquartile ranges [IQR], when applicable) were presented for continuous variables. Categorical variables were described by their absolute and relative frequencies and missing data were excluded from the analyses.

Pearson's chi-square test was applied to assess the association between MCPyV and PD-L1.

Outcomes were evaluated from the date of the first histopathological report confirming the MCC diagnosis and the date of the first recurrence/disease progression or death for progression-free survival (PFS), and patients presenting no recurrence/progression or who died were censored on the date of the last contact. Concerning overall survival (OS), the interval between the histopathological MCC diagnosis date and the date of death from any cause was calculated, and patients alive or lost during follow-up were censored on the date of the last contact. The Kaplan-Meier method was employed to estimate the PFS and OS medians and the curves were compared by the log-rank test [17]. The Hazard Ratios (HR) and the respective confidence intervals (95%CI) for each risk factor were obtained by the semiparametric Cox model [18]. Variables were manually included for multiple model adjustment, according to p < 0.10 values obtained by the univariate Cox model. A *p*-value of < 0.05was considered as statistically significant [18].

## Results

A total of 65 patients were included in the study, 37 males (56.9%) and 28 females (43.1%). Patient baseline demographic, clinical and pathological characteristics are described in Table 1. Mean age at diagnosis was of 72 years old (SD 13.9). Patients were predominantly white (86.2%), with 56.9% (37/64) illiterate or who had not completed elementary school and 41.5% (27/64) completing elementary school or above.

Comorbidities were observed in 41 patients, with systemic arterial hypertension present in 50.8% (33/65) of cases, diabetes mellitus in 23.1% (15/65), chronic renal failure in 6.2% (4/65), HIV infection in 3.1% (2/65) and lymphoproliferative neoplasia in 1.5% (1/65).

The most common primary tumor site was the head and neck region (38.5%), followed by the lower limbs (32.3%), upper limbs (15.4%) and trunk (13.8%).

**Table 1** Baseline
 demographic,
 clinical
 and
 pathological

 characteristics

	n (%)
Age (years)	
Mean (SD)	72 (13.9)
Median (IQR)	73 (19)
Sex	
Male	37 (56.9)
Female	28 (43.1)
Race	
White	56 (86.2)
Other	9 (13.8)
Education	
Illiterate or incomplete elementary school	37 (56.9)
Complete elementary school or above	27 (41.5)
Missing	1 (1.6)
Primary tumor site	
Head and neck	25 (38.5)
Lower limbs	21 (32.3)
Upper limbs	10 (15.4)
Trunk	9 (13.8)
Clinical diameter of the primary tumor in millimeters	
Median (IQR)	40 (45.5)
Disease stage	
-	31 (47.7)
111	13 (20.0)
IV	14 (21.5)
Missing	7 (10.8)
Comorbidities	
Hypertension	33 (50.8)
Diabetes mellitus	15 (23.1)
Chronic renal failure	4 (6.2)
HIV	2 (3.1)
Lymphoproliferative neoplasia	1 (1.5)
Initial treatment	
Surgery	44 (67.7)
Chemotherapy	8 (12.3)
Radiotherapy	17 (26.2)
Immunotherapy	0 (0)
General histopathological aspects	
Mitotic index—Median (IQR)	24/10HPF (37)
Fibroplasia present	59 (90.8)
Tumor thickness in millimetres—Mean (SD)	17.9 (26.5)
Ulceration present	20 (30.8)
Necrosis present	22 (33.8)
Hemorrhage present	31 (47.7)
Histological subtype	
Nodular	40 (61.5)
Infiltrative	24 (36.9)
Missing	1 (1.6)

HPF: high power field. IQR: SD: standard deviation. IQR: Interquartile range

Concerning the clinical primary tumor diameter, the minimum reported size was of 4 mm and the maximum, 150 mm, with a median of 40 mm (IQR 45.5). According to the AJCC 8th edition, 47.7% (31/58) of the cases were classified as stage I or II, 20.0% (13/58) as stage III and 21.5% (14/58) as stage IV.

The mitotic index median was 24/10 high power field (IQR 37). Fibroplasia was present in 90.8% (59/65) of the cases. Mean tumor thickness was 17.9 mm (minimum 1.8 mm and maximum 160 mm). Ulcerations were present in 30.8% (20/65) of the patients, necroses in 33.8% (22/65) and hemorrhages, in 47.7% (31/65). The most frequent subtype was nodular, in 61.5% (40/64) of the cases, while the infiltrative subtype was observed in 36.9% (24/64).

A total of 29 cases were positive for MCPyV (44.6%) (Fig. 1). PD-L1 expression was  $\geq$  1% in 42 cases (64.6%) and  $\geq$  5% in 17 cases (26.2%). Among the 42 patients in which PD-L1 was different from 0%, a focal distribution pattern was observed in 17 cases (40.5%), multifocal in 16 cases (38.1%), diffuse in eight cases (19.0%) and undetermined in one (2.4%) (Fig. 2). Significant associations between MCPyV and PD-L1 expression  $\geq$  1% (p=0.003) and PD-L1 expression  $\geq$  5% (p=0.005) were noted (Tables 2 and 3).

The median follow-up period was of 98.6 months. The estimated PFS at 60 months and 120 months for stages I, II and III were 42.6% (95%CI 30.1–60.3%) and 18% (95%CI 6.6–48.7%), respectively.

Absolute number of deaths comprised 49 cases, 43 due to MCC and six from other causes, namely hemorrhagic stroke, acute myocardial infarction, prostate cancer, skin squamous cell carcinoma, COVID-19, and one external cause.

Concerning the univariate analysis, being male (HR=2.15, 95%CI 1.11 to 4.15, p=0.023), with a low education level (HR=2.68, 95%CI 1.40–5.11, p=0.003) and presenting an advanced MCC stage (HR=4.11, 95%CI 1.95–8.68, p<0.001) were associated with worse PFS. In the multivariate analysis, only education level (HR=2.29, 95%CI 1.19–4.41, p=0.013) and advanced stage (HR=3.36, 95%CI 1.58–7.18, p=0.002) led to statistically significant worse PFS (Table 4).

The estimated OS at 60 months and 120 months for stages I, II and III were 48.9% (95%CI 35.9–66.2%) and 17.8% (95%CI 6.5–48.5%), respectively, and 8.9% for stage IV only at 60 months (95%CI 1.4–56%).

Being male, education level and advanced MCC stage were associated with worse OS in both the univariate and multivariate analysis. In the multivariate



Fig. 1 Immunohistochemistry A positive control for MCPyV (40 × magnification). B MCPyV-positive case (40 × magnification). C) MCPyV-negative case (4× magnification)



Fig. 2 Immunohistochemistry A PD-L1-positive control (40 × magnification). B PD-L1-positive case—80% positivity with a diffuse distribution pattern (10 × magnification)

**Table 2** Pearson's chi-square test with continuity correction(Yates correction) comparing the dichotomous variable PD-L1(1%) by MCPyV status

	PD-L1 < 1%	$PD\text{-}L1 \ge 1\%$	<i>p</i> -value
Negative MCPyV	19 (52.8%)	17 (47.2%)	0.003
Positive MCPyV	4 (13.8%)	25 (86.2%)	
Total	23	42	

**Table 3** Pearson's chi-square test with continuity correction(Yates correction) comparing the dichotomous variable PD-L1(5%) by MCPyV status

	PD-L1 < 5%	PD-L1≥5%	<i>p</i> -value
Negative MCPyV	32 (88.9%)	4 (11.1%)	0.005
Positive MCPyV	16 (55.2%)	13 (44.8%)	
Total	48	17	

analysis, being male (HR = 2.30, 95%CI 1.14–4.67, p = 0.021), education level (HR = 2.13, 95%CI = 1.09–4.19, p = 0.028) and advanced MCC stage (HR = 3.31, 95%CI 1.54–7.09, p = 0.002) led to statistically significant worse OS (Table 5). The OS curves according to the stage, sex and education level are depicted in Fig. 3.

MCPyV was not statistically correlated with PFS (HR = 0.67, 95%CI 0.36–1.25, p = 0.211) or OS (HR = 0.70, 95%CI 0.37–1.33, p = 0.278). Likewise, PD-L1 expression was not statistically correlated with PFS (HR = 0.79, 95%CI 0.43–1.47, p = 0.461) or OS (HR = 0.84, 95%CI 0.45–1.59, p = 0.598) (Table 5).

#### Discussion

MCC rates are usually higher in men (62.1%) compared to women (37.9%) considering most available epidemiological data [14]. In Brazil, Melo et al. published a study on 881 MCC patients reporting a slight female predominance (51.2%) [19]. MCC occurs more frequently in patients over 60 [14, 19, 20]. Herein, MCC was noted as more frequent in male patients, with a mean age of 72, similar to that described in the literature. A white ethnicity is reported as predominant in the literature [14, 19], also noted herein.

Being older and male are associated with worse OS according to the literature [21, 22], although age was not a prognostic factor in the present study.

Herein, low education levels (illiterate or incomplete elementary school) comprised a risk factor concerning PFS and OS in the multivariate analysis. Data correlating schooling and survival in MCC cases, however, are not yet been available. Conversely, correlations between low education and worse survival have been described for other malignant tumors [23, 24]. Low education levels can be associated with lower treatment adherence, greater difficulty in accessing health services, and consequently, late diagnoses, all of which negatively influence case management and outcomes, potentially explaining the obtained results.

According to the National Cancer Database (NCDB) data, MCC is most common in the head and neck region (42.6%), followed by upper limbs and shoulders (23.6%) [14]. Andea et al. reported the extremities as the most common primary tumor site (42%), followed by the head and neck (37%) [25]. In the present study, the most common topography was head and neck, curiously followed by the lower limbs.

Average clinical MCC tumor sizes in the literature range from 7 to 30 mm [26-30]. Herein, however, the

## Table 4 Univariate and multivariate analyses for PFS

	n	HR (univariate)	HR (multivariate)
Sex			
Female	23	_	_
Male	35	2.15 (1.11–4.15, p=0.023)	
Education level			
Complete elementary school or above	26	_	_
Illiterate or incomplete elementary school	32	2.68 (1.40–5.11, p=0.003)	2.29 (1.19–4.41, p=0.013)
Disease stage			
-	44	_	_
IV	14	4.11 (1.95–8.68, <i>p</i> < 0.001)	3.36 (1.58–7.18, p=0.002)
Surgery (initial treatment)			
No	19	_	_
Yes	39	0.25 (0.13–0.47, <i>p</i> < 0.001)	
MCPyV			
Negative	31	_	_
Positive	27	0.67 (0.36–1.25, <i>p</i> =0.211)	
PD-L1 expression			
<1%	21	_	_
≥1%	37	0.79 (0.43–1.47, <i>p</i> =0.461)	

HRs and respective 95%CIs estimated by the Cox model for the semiparametric PFS outcome

#### Table 5 Univariate and multivariate analyses for OS

	n	HR (univariate)	HR (multivariate)
Sex			
Female	23	_	_
Male	35	2.67 (1.33–5.38, <i>p</i> =0.006)	2.30 (1.14–4.67, <i>p</i> =0.021)
Education level			
Complete elementary school or above	26	_	_
Illiterate or incomplete elementary school	32	2.59 (1.33–5.03, <i>p</i> =0.005)	2.13 (1.09–4.19, p=0.028)
Disease stage			
-	44	_	_
IV	14	3.72 (1.77–7.81, p=0.001)	3.31 (1.54–7.09, <i>p</i> =0.002)
Surgery (initial treatment)			
No	19	_	_
Yes	39	0.25 (0.13–0.49, <i>p</i> < 0.001)	
MCPyV			
Negative	31	_	_
Positive	27	0.70 (0.37–1.33, <i>p</i> =0.278)	
PD-L1 expression			
< 1%	21		-
≥1%	37	0.84 (0.45–1.59, <i>p</i> = 0.598)	

HRs and respective 95%Cls estimated by the Cox model for the semiparametric OS outcome

median tumor diameter was larger than published data. Similarly, mean tumor thickness reported in the literature ranges from 8.8 to 12.3 mm [25, 31], with the mean thickness observed in this cohort also larger than literature data. This may be due to delays in accessing the tertiary

site, resulting in large and profound primary tumors at the time of hospital admission.

According to the literature, immunosuppression comprises an MCC risk factor, which is more common in transplanted patients, patients presenting



Fig. 3 A Overall Survival Curve according to AJCC staging 8th edition. B Overall Survival Curve according to sex. C Overall Survival Curve according to education level

lymphoproliferative neoplasms and HIV-infected patients [32, 33]. A low frequency of these comorbidities was, however, observed herein. Systemic arterial hypertension, on the other hand, was present in 50.8% of the cases and diabetes mellitus, in 23.1%, potentially due to the confounding factor of advanced age, as these comorbidities are more common in elderly patients.

Some studies have not reported correlations between MCC thickness and OS [29, 31]. Andea et al. however, in their study on 156 patients, reported that thicker tumors were associated with decreased survival rates [25]. Herein, tumor thickness alone was not correlated to PFS or OS.

Llombart et al. reported a predominant nodular tumor growth pattern in 70% of their cases [29]. Mott et al. reported that an infiltrative growth pattern is associated with adverse prognosis [30], while two studies have demonstrated that a nodular pattern is related to better survival rates [25, 34].

Melo et al., in a study conducted in Brazil, reported that MCC diagnosis occurs predominantly at stages III or IV (50.5%) [19]. Herein, most patients were diagnosed at stages I and II.

According to the AJCC Cancer Staging Manual 8th edition, 5-year OS estimates for clinical staging comprise 45.0% for stage I, 30.9% IIA and 27.3% for stage IIB, respectively. The 5-year OS for the revised stage IIIA was 40.3% and 26.8% for stage IIIB [14]. Considering stage IV, the 2-year survival rate is of only 26% [1]. In the present study, the estimated 5-year OS for stage IV was only lower than the AJCC estimates, probably due to the previously mentioned service access issues and the lack of checkpoint inhibitors for systemic treatment.

The frequency of MCPyV detection depends on the applied method. Busam et al. reported 88% MCPyV positivity using polymerase chain reaction (PCR), decreasing to 67% when applying immunohistochemistry [35]. Similarly, Leroux-Kozal et al. reported 88% and 58% MCPyV positivity when employing PCR and immunohistochemistry, respectively [36]. Paik et al. reported that only 19 out of 104 (18.3%) Australian MCC cases exhibited positive immunohistochemical staining for MCPyV [37]. Another study concerning 37 cases using PCR suggested that MCPyV prevalence may be as low as 24% for Australian patients. The authors postulate that this may be explained due to greater ultraviolet exposure in Australia, resulting in MCC carcinogenesis uncontrolled by MCPyV [38]. Herein, almost half of the cases were MCPvV positive as determined by immunohistochemistry.

Several studies support a more favorable prognosis of MCPyV-positive MCCs. Moshiri et al., for example, in a study involving 282 cases, reported a better prognosis in MCPyV-positive cases. In another study, a multivariate

analysis including age, sex and immunosuppression, indicated that patients with virus-negative MCC exhibited a significantly increased risk of disease progression (HR=1.77, 95%CI 1.20–2.62) and death (HR=1.85, 95%CI 1.19–2.89) [39]. Furthermore, Sihto et al., reported that MCPyV-positive cases display a higher survival rate compared to MCPyV-negative counterparts, presenting a 5-year survival of 45% versus 13% (p < 0.01), respectively [40]. Comparing this to the data obtained in the current cohort, MCPyV positivity exhibited a trend towards a protective exposure profile, albeit non-statistically significant, probably due to the small sample size.

Lipson et al. observed PD-L1 positivity in 49% of 67 MCC samples, considering a cutoff point of 5%. These authors also reported a statistically significant association between tumor cell PD-L1 expression and longer survival times (HR=3.12, 95%CI 1.28–7.61, p=0.012) [41]. Herein, the number of PD-L1-positive cases was lower when the cutoff was set at 5%. No statistically significant association was detected in the univariate analysis concerning the OS and PFS analysis, even when considering a cut-off point of 1% for PD-L1 positivity.

A significant association has been noted between MCPyV and PD-L1 in MCC tumors [41], with other studies observing similar results [42–44]. In this regard, MCPyV infection seems to promote the expression of immune response-associated proteins [42]. In the present study, a significant association between MCPyV and PD-L1 expression was observed.

Some limitations should be acknowledged concerning this assessment. First, the fact that this is retrospective study, which makes data collection difficult. It is also a single-center study encompassing a limited sample size, posing difficulties in conducting robust statistical analyses and increasing the margin of error. The rarity of the disease is also noteworthy, hindering cohort studies and leading to prolonged intervals concerning case selections. Moreover, during this period, the review and updating of MCC treatment protocols during the study period make it challenging to verify treatment efficacy. Finally, a considerable limitation arises from the unavailability if immunotherapy at the Brazilian National Cancer Institute, where the research was conducted, which may potentially impact survival data when compared to other institutions that offer this contemporary treatment.

## Conclusions

This is the first large Brazilian cohort to assess the prevalence of MCPyV in tumors, as well as PD-L1 expression and the association between these markers. The prevalence of MCPyV infection in MCC cases in the studied population was of 44.6%, although no statistically significant correlations with PFS and OS

were observed. PD-L1 expression was  $\geq 1\%$  in 42 cases (64.6%) and  $\geq 5\%$  in 17 cases (26.2%). No statistical significance concerning PD-L1 correlations to PFS and OS were, however, noted. Significant associations between MCPyV and PD-L1 expression  $\geq 1\%$  (p = 0.003) and PD-L1 expression  $\geq 5\%$  (p = 0.005) were detected.

#### Abbreviations

AJCC	American Joint Committee on Cancer
CI	Confidence interval
HR	Hazard ratios
INCA	Brazilian National Cancer Institute
IQR	Interquartile range
MCC	Merkel cell carcinoma
MCPyV	Merkel cell polyomavirus
NCDB	National Cancer Database
OS	Overall survival
PCR	Polymerase chain reaction
PD-1	Programmed cell death 1
PD-L1	Programmed cell death protein ligand 1
PFS	Progression-free survival
REDCap	Research data capture
SD	Standard deviation

#### Acknowledgements

Not applicable.

#### Author contributions

AM and SMS contributed to the study's conception and design. The first draft of the manuscript was written by SMS and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript. The idea for the project and the article came from ACM. The literature search was performed by SMS, GCC, PRGS and ACM. Data analysis was performed by Isabele ÁS. The work was critically revised by ACM.

#### Funding

Funding for this research come from the Division of Clinical Research and Technological Development of Brazilian National Cancer Institute.

#### Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

#### Declarations

#### Ethics approval and consent to participate

All methods were carried out in accordance with relevant guidelines and regulations.

#### **Consent for publication**

Not applicable.

#### Informed consent

Informed consent was obtained from all subjects and/or their legal guardian(s).

#### Human or animal rights

The study was approved by the Ethics in Human Research Committee of the Brazilian National Cancer Institute (Certificate of Presentation of Ethical Appreciation—CAAE: 25426419.2.0000.5274) and conducted following the Good Clinical Practice guidelines.

#### **Competing interests**

The authors declare that they have no competing interests.

#### Author details

<sup>1</sup> Division of Clinical Research and Technological Development, Brazilian National Cancer Institute, Rio de Janeiro, Brazil. <sup>2</sup>Dermatology, Gávea Medical Center, Rio de Janeiro, Brazil. <sup>3</sup>Division of Pathology, Brazilian National Cancer Institute, Rio de Janeiro, Brazil.

Received: 19 September 2023 Accepted: 20 December 2023 Published online: 04 January 2024

#### References

- Wehkamp U, Stern S, Krüger S, Weichenthal M, Hauschild A, Röcken C, Egberts F. Co-expression of NGF and PD-L1 on tumor-associated immune cells in the microenvironment of Merkel cell carcinoma. J Cancer Res Clin Oncol. 2018;144(7):1301–8. https://doi.org/10.1007/s00432-018-2657-x.
- 2. Toker C. Trabecular carcinoma of the skin. Arch Dermatol. 1972;105(1):107–10.
- Harold A, Amako Y, Hachisuka J, Bai Y, Li MY, Kubat L, Gravemeyer J, Franks J, Gibbs JR, Park HJ, Ezhkova E, Becker JC, Shuda M. Conversion of Sox2-dependent Merkel cell carcinoma to a differentiated neuronlike phenotype by T antigen inhibition. Proc Natl Acad Sci U S A. 2019;116(40):20104–14. https://doi.org/10.1073/pnas.1907154116.
- Mazziotta C, Cervellera CF, Badiale G, Vitali I, Touzé A, Tognon M, Martini F, Rotondo JC. Distinct retinoic gene signatures discriminate Merkel cell polyomavirus-positive from -negative Merkel cell carcinoma cells. J Med Virol. 2023;95(7): e28949. https://doi.org/10.1002/jmv.28949.
- Thibault K. Evidence of an epithelial origin of Merkel cell carcinoma. Mod Pathol. 2022;35(4):446–8. https://doi.org/10.1038/s41379-021-00964-x.
- Feng H, Shuda M, Chang Y, Moore PS. Clonal integration of a polyomavirus in human Merkel cell carcinoma. Science. 2008;319(5866):1096–100. https://doi.org/10.1126/science.1152586.
- Harms PW, Harms KL, Moore PS, et al. The biology and treatment of Merkel cell carcinoma: current understanding and research priorities. Nat Rev Clin Oncol. 2018;15(12):763–76. https://doi.org/10.1038/ s41571-018-0103-2.
- 8. DeCaprio JA. Molecular pathogenesis of Merkel cell carcinoma. Annu Rev Pathol. 2021;16:69–91. https://doi.org/10.1146/annurev-pathm echdis-012419-032817.
- Mazziotta C, Lanzillotti C, Gafà R, Touzé A, Durand MA, Martini F, Rotondo JC. The role of histone post-translational modifications in merkel cell carcinoma. Front Oncol. 2022;12: 832047. https://doi.org/10.3389/fonc. 2022.832047.
- Cook M, Baker K, Redman M, Lachance K, Nguyen MH, Parvathaneni U, Bhatia S, Nghiem P, Tseng YD. Differential outcomes among immunosuppressed patients with Merkel cell carcinoma: impact of immunosuppression type on cancer-specific and overall survival. Am J Clin Oncol. 2019;42(1):82–8. https://doi.org/10.1097/COC.000000000000482.
- 11. Coursaget P, Samimi M, Nicol JT, Gardair C, Touzé A. Human Merkel cell polyomavirus: virological background and clinical implications. APMIS. 2013;121(8):755–69. https://doi.org/10.1111/apm.12122.
- McEvoy AM, Lachance K, Hippe DS, Cahill K, Moshiri Y, Lewis CW, Singh N, Park SY, Thuesmunn Z, Cook MM, Alexander NA, Zawacki L, Thomas H, Paulson KG, Nghiem P. Recurrence and mortality risk of Merkel cell carcinoma by cancer stage and time from diagnosis. JAMA Dermatol. 2022;158(4):382–9. https://doi.org/10.1001/jamadermatol.2021.6096.
- Samimi M. Immune checkpoint inhibitors and beyond: An overview of immune-based therapies in merkel cell carcinoma. Am J Clin Dermatol. 2019;20(3):391–407. https://doi.org/10.1007/s40257-019-00427-9.
- Harms KL, Healy MA, Nghiem P, Sober AJ, Johnson TM, Bichakjian CK, Wong SL. Analysis of prognostic factors from 9387 Merkel cell carcinoma cases forms the basis for the new 8th edition AJCC staging system. Ann Surg Oncol. 2016;23(11):3564–71. https://doi.org/10.1245/ s10434-016-5266-4.
- Smoller BR, Bichakjian C, Brown JA, Crowson AN, Divaris D, Frishberg DP, Gao L, Gershenwald J, McNiff JM, Nghiem P, Prieto VG, Scolyer RA, BMed-Sci, Selim MA, Shalin SC, Taube JM (2021) Protocol for the examination of specimens from patients with Merkel cell carcinoma of the skin, Version: 4.1.0.0. College of American Pathologists Cancer Protocol Templates

- R Core Team (2022) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL https:// www.R-project.org/
- Clark TG, Bradburn MJ, Love SB, Altman DG. Survival analysis part I: basic concepts and first analyses. Br J Cancer. 2003;89(2):232–8. https://doi.org/ 10.1038/sj.bjc.6601118.
- Bradburn MJ, Clark TG, Love SB, Altman DG. Survival analysis part II: multivariate data analysis–an introduction to concepts and methods. Br J Cancer. 2003;89(3):431–6. https://doi.org/10.1038/sj.bjc.6601119.
- Melo AC, Thuler LS. Trends in the incidence and morbidity of Merkel cell carcinoma in Brazil. Future Oncol. 2020;17(22):2857–65. https://doi.org/ 10.2217/fon-2020-1313.
- Youlden DR, Soyer HP, Youl PH, Fritschi L, Baade PD. Incidence and survival for Merkel cell carcinoma in Queensland, Australia, 1993–2010. JAMA Dermatol. 2014;150(8):864–72. https://doi.org/10.1001/jamadermat ol.2014.124.
- Chen MM, Roman SA, Sosa JA, Judson BL. The role of adjuvant therapy in the management of head and neck Merkel cell carcinoma: an analysis of 4815 patients. JAMA Otolaryngol Head Neck Surg. 2015;141(2):137–41. https://doi.org/10.1001/jamaoto.2014.3052. (PMID: 25474617).
- Dudzisz-Sledz M, Sobczuk P, Kozak K, Switaj T, Kosela-Paterczyk H, Czarnecka AM, Falkowski S, Rogala P, Morysinski T, Spalek MJ, Zdzienicki M, Goryn T, Zietek M, Cybulska-Stopa B, Klek S, Kaminska-Winciorek G, Ziolkowska B, Szumera-Cieckiewicz A, Rutkowski P. Treatment of locally advanced Merkel cell carcinoma-a multi-center study. Cancers (Basel). 2022;14(2):422. https://doi.org/10.3390/cancers14020422.
- Magalhaes LP, Oshima CT, Souza LG, Lima JM, Carvalho L, Forones NM. Variação de peso, grau de escolaridade, saneamento básico, etilismo, tabagismo e hábito alimentar pregresso em pacientes com cancêr de estômago. Arq Gastroenterol. 2008;45(2):111–6.
- Melo WA, Pelloso SM, Alvarenga A, Carvalho MD. Fatores associados a alterações do exame citopatológico cérvico-uterino no Sul do Brasil. Rev Bras Saúde Matern Infant. 2017;17(4):645–52. https://doi.org/10.1590/ 1806-93042017000400002.
- Andea AA, Coit DG, Amin B, Busam KJ. Merkel cell carcinoma: histologic features and prognosis. Cancer. 2008;113(9):2549–58. https://doi.org/10. 1002/cncr.23874.
- Allen PJ, Bowne WB, Jaques DP, Brennan MF, Busam K, Coit DG. Merkel cell carcinoma: prognosis and treatment of patients from a single institution. J Clin Oncol. 2005;23(10):2300–9. https://doi.org/10.1200/JCO.2005.02. 329.
- Allen PJ, Busam K, Hill AD, Stojadinovic A, Coit DG. Immunohistochemical analysis of sentinel lymph nodes from patients with Merkel cell carcinoma. Cancer. 2001;92(6):1650–5.
- Koljonen V, Böhling T, Granhroth G, Tukiainen E. Merkel cell carcinoma: a clinicopathological study of 34 patients. Eur J Surg Oncol. 2003;29(7):607– 10. https://doi.org/10.1016/s0748-7983(03)00110-0.
- Llombart B, Monteagudo C, López-Guerrero JA, Carda C, Jorda E, Sanmartín O, Almenar S, Molina I, Martín JM, Llombart-Bosch A. Clinicopathological and immunohistochemical analysis of 20 cases of Merkel cell carcinoma in search of prognostic markers. Histopathology. 2005;46(6):622–34. https://doi.org/10.1111/j.1365-2559.2005.02158.x.
- Mott RT, Smoller BR, Morgan MB. Merkel cell carcinoma: a clinicopathologic study with prognostic implications. J Cutan Pathol. 2004;31(3):217– 23. https://doi.org/10.1111/j.0303-6987.2004.00149.x.
- Sandel HD 4th, Day T, Richardson MS, Scarlett M, Gutman KA. Merkel cell carcinoma: Does tumor size or depth of invasion correlate with recurrence, metastasis, or patient survival? Laryngoscope. 2006;116(5):791–5. https://doi.org/10.1097/01.mlg.0000208615.93883.b2.
- Bichakjian CK, Olencki T, Aasi SZ, et al. Merkel cell carcinoma, version 1.2018, NCCN clinical practice guidelines in oncology. J Natl Compr Cancer Netw JNCCN. 2018;16(6):742–74. https://doi.org/10.6004/jnccn.2018. 0055.
- Schmults CD, Blitzblau R, Aasi SZ, et al (2019) NCCN guidelines index table of contents discussion. Merkel Cell Carcinoma 66
- Smith FO, Yue B, Marzban SS, Walls BL, Carr M, Jackson RS, Puleo CA, Padhya T, Cruse CW, Gonzalez RJ, Sarnaik AA, Schell MJ, DeConti RC, Messina JL, Sondak VK, Zager JS. Both tumor depth and diameter are predictive of sentinel lymph node status and survival in Merkel cell carcinoma. Cancer. 2015;121(18):3252–60. https://doi.org/10.1002/cncr.29452.

- Busam KJ, Jungbluth AA, Rekthman N, Coit D, Pulitzer M, Bini J, Arora R, Hanson NC, Tassello JA, Frosina D, Moore P, Chang Y. Merkel cell polyomavirus expression in Merkel cell carcinomas and its absence in combined tumors and pulmonary neuroendocrine carcinomas. Am J Surg Pathol. 2009;33(9):1378–85. https://doi.org/10.1097/PAS.0b013e3181aa30a5.
- Leroux-Kozal V, Lévêque N, Brodard V, Lesage C, Dudez O, Makeieff M, Kanagaratnam L, Diebold MD. Merkel cell carcinoma: histopathologic and prognostic features according to the immunohistochemical expression of Merkel cell polyomavirus large T antigen correlated with viral load. Hum Pathol. 2015;46(3):443–53. https://doi.org/10.1016/j.humpath.2014. 12.001.
- Paik JY, Hall G, Clarkson A, Lee L, Toon C, Colebatch A, Chou A, Gill AJ. Immunohistochemistry for Merkel cell polyomavirus is highly specific but not sensitive for the diagnosis of Merkel cell carcinoma in the Australian population. Hum Pathol. 2011;42(10):1385–90. https://doi.org/10.1016/j. humpath.2010.12.013.
- Garneski KM, Warcola AH, Feng Q, Kiviat NB, Leonard JH, Nghiem P. Merkel cell polyomavirus is more frequently present in North American than Australian Merkel cell carcinoma tumors. J Invest Dermatol. 2009;129(1):246–8. https://doi.org/10.1038/jid.2008.229.
- Moshiri AS, Doumani R, Yelistratova L, Blom A, Lachance K, Shinohara MM, Delaney M, Chang O, McArdle S, Thomas H, Asgari MM, Huang ML, Schwartz SM, Nghiem P. Polyomavirus-negative merkel cell carcinoma: a more aggressive subtype based on analysis of 282 cases using multimodal tumor virus detection. J Invest Dermatol. 2017;137(4):819–27. https://doi.org/10.1016/j.jid.2016.10.028.
- Sihto H, Kukko H, Koljonen V, Sankila R, Böhling T, Joensuu H. Clinical factors associated with Merkel cell polyomavirus infection in Merkel cell carcinoma. J Natl Cancer Inst. 2009;101(13):938–45. https://doi.org/10. 1093/jnci/djp139.
- Lipson EJ, Vincent JG, Loyo M, Kagohara LT, Luber BS, Wang H, Xu H, Nayar SK, Wang TS, Sidransky D, Anders RA, Topalian SL, Taube JM. PD-L1 expression in the Merkel cell carcinoma microenvironment: association with inflammation, Merkel cell polyomavirus and overall survival. Cancer Immunol Res. 2013;1(1):54–63. https://doi.org/10.1158/2326-6066. CIR-13-0034.
- 42. Donizy P, Wróblewska JP, Dias-Santagata D, Woznica K, Biecek P, Mochel MC, Wu CL, Kopczynski J, Pieniazek M, Ryś J, Marszalek A, Hoang MP. Merkel cell carcinoma of unknown primary: immunohistochemical and molecular analyses reveal distinct UV-signature/MCPyV-negative and high immunogenicity/MCPyV-positive profiles. Cancers (Basel). 2021;13(7):1621. https://doi.org/10.3390/cancers13071621.
- Ricci C, Righi A, Ambrosi F, Gibertoni D, Maletta F, Uccella S, Sessa F, Asioli S, Pellilli M, Maragliano R, La Rosa S, Papotti MG, Asioli S. Prognostic impact of MCPyV and TIL subtyping in merkel cell carcinoma: evidence from a large european cohort of 95 patients. Endocr Pathol. 2020;31(1):21–32. https://doi.org/10.1007/s12022-019-09601-5.
- Walsh NM, Castonguay MC, Carter MD, Pasternak S, Ly TY, Doucette S, Hanly JG, Saggini A, Cerroni L. Global PD-L1 signals and tumor-infiltrating lymphocytes: markers of immunogenicity in different subsets of Merkel cell carcinoma and potential therapeutic implications. Am J Dermatopathol. 2019;41(11):819–25. https://doi.org/10.1097/DAD.000000000 001390.

## **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.